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Influence of priming on germination of khirni seedlings (*Manilkara hexandra* L.)

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Abstract

The field trial was conducted at the Nursery and Research Farm, Department of Horticulture, College of Agriculture, Dhule during 2021-22, to study the influence of priming on germination of khirni seedlings (*Manilkara hexandra* L.). The experiment was laid out in a Complete Randomized Block Design with ten treatments viz., T₁ (GA₃ @ 50ppm), T₂ (GA₃ @ 75ppm), T₃ (GA₃ @ 100ppm), T₄ (GA₃ @ 200ppm), T₅ (NAA @ 100ppm), T₆ (KNO₃ @ 1%), T₇ (Cow urine @ 10%), T₈ (Cow urine 100%), T₉ (Cow dung slurry) and T₁₀ Control (Distilled Water) with three replications. The treatment T₄ (GA₃ @ 200ppm) resulted, the minimum days required for germination (30.33), however maximum number of seedlings germinate (42.33) at 15 days after sowing and germination percentage (84.67%) at 15 days after sowing observed in the treatment T₉ (cow dung slurry). Hence, it can be concluded that, the cow dung slurry were beneficial for increasing germination parameters as compared to other treatments under Khandesh region of Maharashtra.

Keywords: Seedlings, germination, soaking treatments, khirni

Introduction

Khirmi (*Manilkara hexandra* Roxb.) commonly known as “Rayan,” or “Ranjana.” The South East Asia was native place of Khirmi in central and peninsular India. Tropical fruit crop grown commercially in India. The tree is about 15-18 metre tall with grey bark, glabrous branchlets, leaves 5-13 cm long, shiny, alternate leaves that are sometimes crowded together at the ends of branchlets and prominent scars. (Sanjay *et al.*, 2017) [22]. Mostly seeds are used to propagate khirmi. It is a slow-growing plant that can withstand drought. Fruits of khirmi have high economical value as mature fresh fruits which are sweet and a good source of iron, minerals, sugars, protein, carbohydrate and vitamin A (Pareek *et al.*, 1998 and Singh *et al.*, 2006) [16, 26]. The different parts of this plant find their use in treatment of ulcers, dyspepsia, opacity of the cornea, bronchitis, urethrorrhea, leprosy, etc. (Anonymous, 1962; Pareek *et al.* 1998; Hoareau, 1999; Raju and Reddy 2005; Chanda and Parekh 2010) [2, 16, 8, 20]. The seeds contain approximately 25% oil, which is used for cooking purpose. The bark also contains 10% tannin, which is used for treatment of fever and may be utilized in tanning purpose (Anonymous, 1962) [2].

Propagation through seeds is considered to be one of the most reliable, efficient and universally applied method for raising seedlings (Hartmann *et al.*, 1990) [6]. The growing medium influences the quality of seedlings produced in a nursery, which in turn affects the re-establishment in the main field (Agbo and Omaliko, 2006) [1]. The influence of the medium is felt even before the plant sprouts because of its water retention and aeration properties. Media composition used as a source of plant nutrients influences the quality of seedling to a considerable extent (Wilson *et al.*, 2001) [29]. It directly affects the development and maintenance of the extensive functional root system. Proper media management is imperative to the production of quality seedlings since vigorous growth is required to face the seasonal hazards encountered on the main field (Khan *et al.*, 2006) [11]. Khirmi seeds have a hard seed coat and are recalcitrant in nature, therefore utmost care has to be taken for enhancing its germination percentage (Samir *et al.*, 2015) [21].

The quality of seedlings obtained from a nursery affects their establishment in the main field and the ultimate productivity of an orchard. Various pre-sowing seed treatments to improve germination and to reduce germination time have been widely investigated in tree species (Prasad and Prasad 2009, Prasad *et al.* 2011) [17, 18, 19]. Several efforts like treatment with chemicals, growth regulators, hot water, cattle urine and cow dung slurry have been used to overcome hard seed coat dormancy.

Use of plant growth regulators in enhancing seedling growth of numerous plant species is well known (Marler and Mickelbart, 1992 and Hazrat *et al.*, 2006) [13, 7]. Khirni seeds have very weak germination rates, and their following growth is also quite little. Khirni seedlings need a lot of time to grow to the right height and vigour for grafting. Improvements have already been made to the khirni seeds germination and subsequent development. It is generally known that bioregulators can improve seed germination and seedling growth in a variety of plant species (Malshe *et al.*, 2014) [12]. In addition, it is known that synthetic chemicals (Vachhani *et al.*, 2014; Jadhav *et al.* 2015) [28, 10] and other naturally occurring bio-products of organics (cow-dung, cow urine) contain essential plant growth substances that promote plant growth and development (Anonymous, 1993; Shirol *et al.*, 2005; Shinde and Malshe, 2015) [3, 25, 24]. To study the impact of various chemicals, plant growth regulators, cow dung slurry, and cow urine on the germination, growth, and development of khirni seedlings.

Materials and Methods

An experiment on khirni seeds was conducted at the Nursery and Research Farm, Department of Horticulture, College of Agriculture, Dhule during the year 2021-22. The experiment was laid out in a Complete Randomized Block Design with ten treatments *viz.*, T₁ (GA₃ @ 50ppm), T₂ (GA₃ @ 75 ppm), T₃ (GA₃ @ 100 ppm), T₄ (GA₃ @ 200ppm), T₅ (NAA @ 100 ppm), T₆ (KNO₃ @ 1%), T₇

(Cow urine @ 10%), T₈ (Cow urine 100%), T₉ (Cow dung slurry) and T₁₀ Control (Distilled Water) with three replications under 50% shade net condition. The sowing was done in black polythene bags (8'' x 6'') filled with soil and FYM (3:1) in July, 2021. The statistical analysis of the data in respect of germination was done according to the standard procedure given by Panse and Sukhatme (1984) [15].

Result and Discussion

Influence of priming on germination parameters of khirni seedlings

It is revealed from the data (Table 1 and Fig. 1-3), the significantly minimum days required for germination (30.33) recorded in treatment T₄ (GA₃ @ 200ppm) which was at par with the treatment T₃ (GA₃ @ 100ppm) 31.33, T₉ (Cow dung slurry) 31.67, T₈ (Cow urine @ 100%) 32.00, T₅ (NAA @ 100ppm) 32.33, T₂ (GA₃ @ 75ppm) 33.00 and T₇ (Cow urine @ 10%) 33.33 However, the maximum number of seedlings germinate (42.33) recorded in treatment T₉ (Cow dung slurry) which was at par with the treatment, T₈ (Cow urine @ 100%) 41.67, T₄ (GA₃ @ 200ppm) 40.33 and T₃ (GA₃ @ 100ppm) 39.33. at 15 days after sowing and germination percentage (84.67%) at 15 days after sowing recorded in the treatment T₉ (cow dung slurry) which was statistically at par with the treatment, T₈ (Cow urine @ 100%) 83.33%, T₄ (GA₃ @ 200ppm) 80.67% and T₃ (GA₃ @ 100ppm) 78.67%.

Table 1: Influence of priming on germination parameters of khirni seedlings.

Treatments		Days required for germination	Numbers of seedling germinate at 15 th DAS	Germination percentage (%) at 15 th DAS
T ₁	GA ₃ @ 50ppm	36.33	36.00	72.00
T ₂	GA ₃ @ 75ppm	33.00	37.67	75.33
T ₃	GA ₃ @ 100ppm	31.33	39.33	78.67
T ₄	GA ₃ @ 200ppm	30.33	40.33	80.67
T ₅	NAA @ 100ppm	32.33	38.00	76.00
T ₆	KNO ₃ @ 1%	35.00	32.67	65.33
T ₇	Cow urine @ 10%	33.33	36.33	72.67
T ₈	Cow urine @ 100%	32.00	41.67	83.33
T ₉	Cow dung slurry	31.67	42.33	84.67
T ₁₀	Control (Distilled water)	46.00	29.67	59.33
S.E.±		1.02	1.09	2.18
C.D at 5%		3.00	3.22	6.43

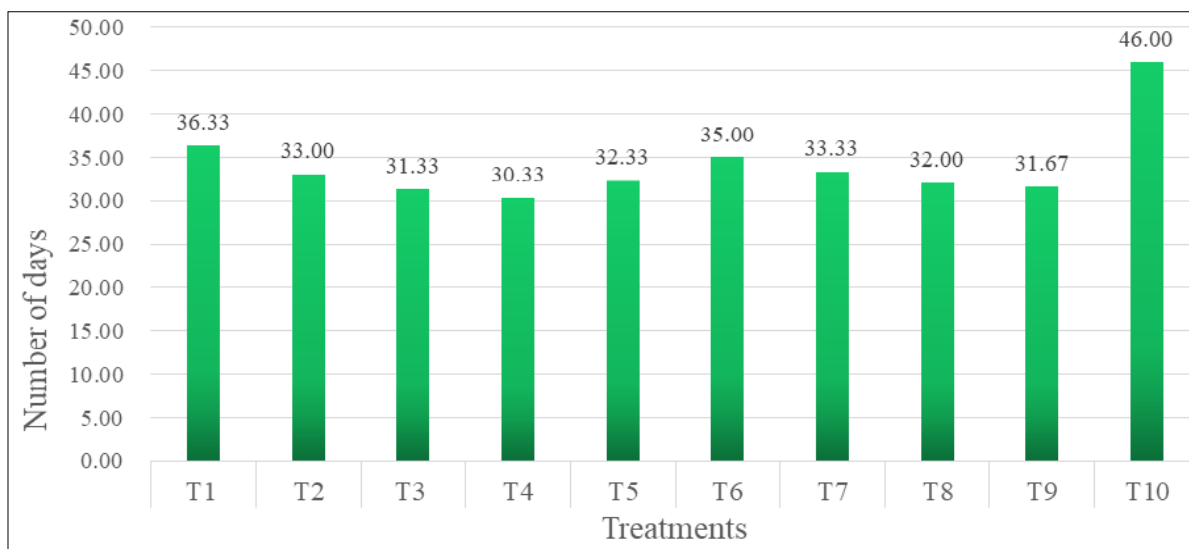


Fig 1: Effect of priming on days required for initiation of germination of khirni.

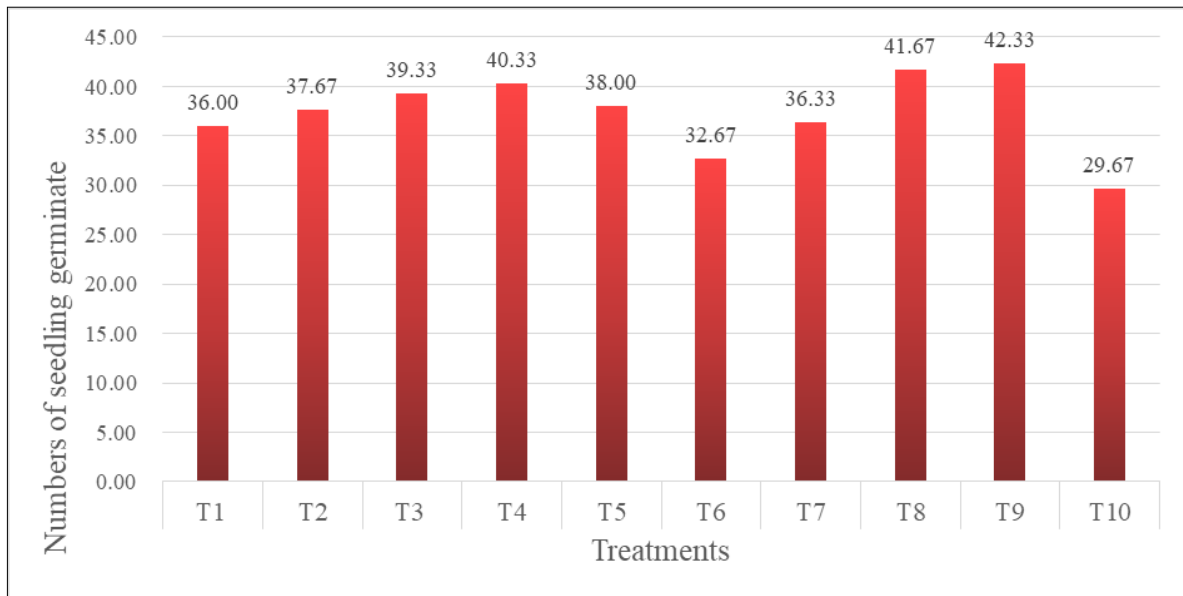


Fig 2: Effect of priming on numbers of seedling germinate of khirni.

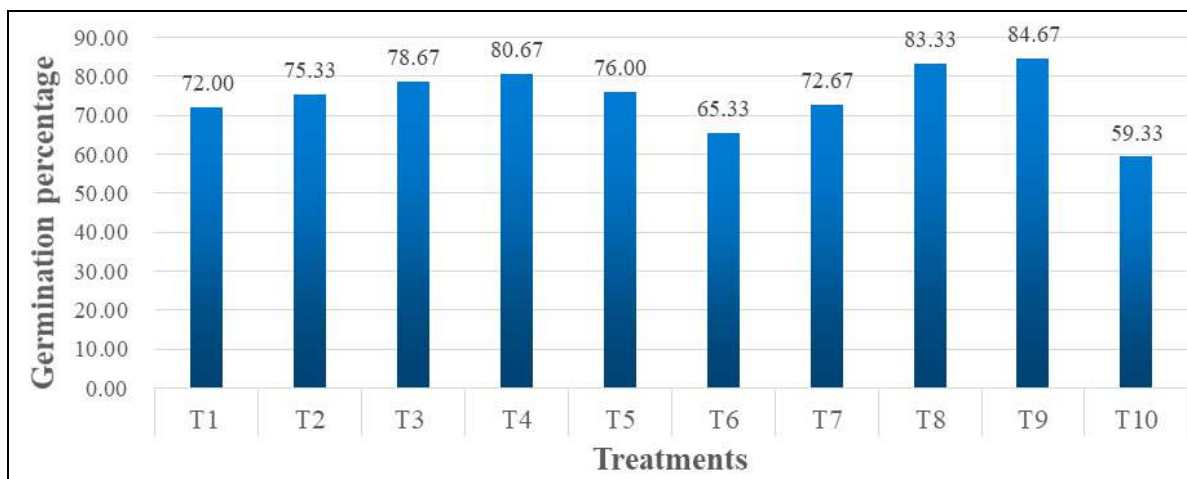


Fig 3: Effect of priming on germination percentage of khirni.

The minimum days required for initiation of germination in treatment GA₃ might be due to the seed treatment with gibberellic acid (GA₃) control mobilization of starch which acts as a respiratory substrate leading to immediate enhancement in cell elongation (Shah, 2007) [23]. GA₃ regulator helps to increase the activities of hydrolyzing enzymes and alpha amylase at initial stage of seed germination and thus facilitated the germination process. These findings are in accordance with the results obtained by Singh *et al.* (1979) [27] in citrus, Hore and Sen (1994) [9] in ber, Pampanna and Sulikeri (2001) [14] and Desai *et al.* (2017) [5] in papaya. The higher number of seedling germinate and germination percentage in cow dung slurry treatment may be attributed to the presence of growth promoting substances (auxins) and NPK nutrients in cattle cow dung. These results are in agreement with the findings of Shirol *et al.* (2005) [25] in khirni rootstock.

Conclusion

The treatment cow dung slurry were found beneficial for increasing the number of seedling germination and percentage in khirni seedlings under Khandesh region of Maharashtra.

References

1. Agbo CU, Omaliko CM. Initiation and growth of shoots of *Gongronema latifolia* Benth stem cuttings in different rooting media. *African J. Biotechnology*. 2006;5:425-428.
2. Anonymous. The wealth of India: raw materials, vol 6. Publications and Information Directorate, CSIR, New Delhi; c1962. p. 298-300.
3. Anonymous. Research Report on Tropical Fruits. Proceedings of Group Discussion of the All India Co-ordinated Research Project on Tropical Fruits, Technical Document No. 58, Indian Institute of Horticultural Research, Bangalore; c1993, p. 143-145.
4. Chanda S, Parekh J. Assessment of antimicrobial potential of *Manilkara hexandra* leaf. *Phcog. J.* 2010;2(12):448-455.
5. Desai A, Panchal B, Trivedi A, Prajapati D. Studies on seeds germination and seedling growth of papaya (*Carica papaya* L.) cv. Madhubindu as influenced by media, GA₃ and cow urine under net 76 house condition. *J. Pharmacognosy and phytochemistry*. 2017;6(4):1448-1451.
6. Hartman HT, Kester DE, Davies FT. *Plant propagation: Principles and Practices*. 5th Edition. Englewood cliffs.

- New Jersey, Prentice Hall; c1990, p.100-250.
7. Hazrat G, Abdul MK, Noorul A. Accelerating the growth of *Araucaria heterophylla* seedlings through different gibberellic acid concentrations and nitrogen levels. *J Agril. Bio. Sci.* 2006;1(2): 25-29.
 8. Hoareau L. Medicinal plants: a re-emerging health aid. *Electron. J Biotechnol.* 1999;2(2):56-70.
 9. Hore JK, Sen SK. Role of pre-sowing seed treatment on germination, seedling growth and longevity of ber (*Ziziphus mauritiana* L.) seeds. *Indian J. of Agricultural Research.* 1994;28(4):284-285.
 10. Jadhav, Archana C, Bhagure YL, Rajeshwari M, Raundal. Effect of Plant Growth Regulator, chemicals and plant extract on seed germination and seedling growth of custard apple (*Annona squamosa*). *Asian Journal of Horticulture.* 2015;10(1):184-186.
 11. Khan MM, Khan MA, Mazhar A, Muhammad J, Ali JMA, Abbas H. Evaluation of potting media for the production of rough lemon nursery stock. *Pakistan J. Botany.* 2006;38(3):623-629.
 12. Malshe KV, Mahadik SG, Desai BG, Borate HV. Effect of pre sowing treatment with Gibberellic acid on germination and 78 growth of khirni (*Manilkara hexandra* L.). *Annual Plant Physiology.* 2014;28(2):73-75.
 13. Marler TE, Mickelbart MV. Application of GA4+7 to stem enhances the carambola seedling growth. *Hort. Sci.* 1992;27(2):122-123.
 14. Pampanna Y, Sulikeri GS. Effect of growth regulators on seed germination and seedling growth of sapota. *Karnataka J of Agric. Sci.* 2001;14(4):1030-1036.
 15. Panse VS, Sukhatme PV. *Statistical Methods for Agricultural Workers*, ICAR, New Delhi; c1984.
 16. Pareek OP, Sharma S, Arora RK. Underutilized edible fruits and nuts: an inventory of genetic resources in their regions of diversity. IPGRI office for South Asia, New Delhi; c1998.
 17. Prasad B, Prasad R. Influence of growth regulators on seed germination and seedling vigour in West Himalayan banj oak (*Quercus leucotrichophora* A. Camus). *Seed Research.* 2009;37(1-2):76-80.
 18. Prasad B, Prasad R. Selection of suitable growth regulators and its concentration for better germination and seedling growth of Himalayan Dogwood (*Benthamidia capitata* Wall ex. Roxb.). *Indian Journal of Forestry.* 2009;32(4):523-527.
 19. Prasad B, Prasad R, Sah VK. Pre-sowing seed treatment of bioagents on seed germination and seedling vigour of Banj oak (*Quercus leucotrichophora* A. Camus). *Indian Journal of Forestry.* 2011;34(1):71-74.
 20. Raju VS, Reddy KN. Ethno-medicine for dysentery and diarrhea from Khammam district of Andhra Pradesh. *Ind. J Trad. Knowl.* 2005;4(4):443-447.
 21. Samir M, Rai R, Prasad B. Seed germination behaviour as influenced by pre-sowing treatments in khirni. *J. Hill Agriculture.* 2015;6(1):133-136.
 22. Sanjay Singh, Singh AK, Apprao VV, Bharagava R. Genetic Divergence in Khirni (*Manilkara Hexandra*) under Semi- Arid Ecosystem of Western India. *The Indian Journal of Agricultural Sciences.* 2017;87(3):616-617.
 23. Shah SH. Physiological effect of pre-sowing seed treatment with gibberellic acid on *Nigella sativa* L. *Acta Botanica Croatica.* 2007;66(1):67-73.
 24. Shinde VV, Malshe KV. Effect of cattle urine and cow dung slurry as seed treatment on germination and growth of (*Manilkara hexandra* L.). *J Ecofriendly Agric.* 2015;10(2):128-130.
 25. Shirol AM, Hanamashetti SI, Kanamadi VC, Thammaiah N, Patil S. Studies on Pre-Soaking, Method and Season of Grafting of Sapota Rootstock Khirnee. *Karnataka J of Agric. Sci.* 2005;18(1):96-100.
 26. Singh Sanjay, Singh AK, Joshi HK, Bagle BG, Dhandar DG. Genetic diversity in Rayan (*Manilkara hexandra*) under semi arid tropics of western India. National symposium on production, utilization and export of underutilized fruits with commercial potentialities, 22-24th November, 2006 at BCKV, Kalyani, West Bengal; c2006. p. 10.
 27. Singh HK, Shankar G, Makhija MA. A study on citrus seed germination as affected by some chemicals. *Haryana J. of Horticultural Sci.* 1979;8(3-4):194-195.
 28. Vachhani KB, Ray NR, Sitapara HH, Patel AP, Mahida SV. Influence of chemicals, Plant Growth Regulators and cow dung slurry as seed treatment on germinability, growth and development of khirni (*Manilkara hexandra* Roxb). *Proceeding Global Conference on Technological Challenges and Human Resources for Climate Smart Horticulture - Issues and Strategies.* May 28-31, 2014, Navsari Agricultural University, Navsari, Gujarat; c2014. p.175.
 29. Wilson SB, Stoffella PJ, Graetz DA. Use of compost as a media amendment for containerized production of two subtropical perennials. *J Envir. Horticulture.* 2001;19:37-42.